

# DNP enhanced frequency-selective TEDOR experiments in bacteriorhodopsin

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# Objectives

- To introduce a new approach (FS-TEDOR) to multiple  $^{13}\text{C}$ - $^{15}\text{N}$  distances measurements in uniformly labeled solids.
- To increase sensitivity by integrating high field DNP to the experiments.
- To demonstrate the method as a study tool for  $^{15}\text{N}$ - $^{13}\text{C}$  correlation spectroscopy in crystalline solids and membrane proteins
- To resolve correlation spectrum of Arg,  $^{13}\text{C}_\gamma$ - $^{15}\text{N}_\epsilon$  region in [U- $^{13}\text{C}$ ,  $^{15}\text{N}$ ] - bacteriorhodopsin

# Introduction

**Heteronuclear distance measurements** in **uniformly labeled sample** under MAS-NMR include complications:

- **Strong dipolar coupling**

- The coupling dominates the spin dynamics and compromises sensitivity
- The effect is crucial in protein with amino acids containing nitrogen side chain ( Asp, Gln, Lys, Arg)

**Solution: Frequency selective to solve N in backbone or sidechain**

- **J-coupling ( $^{13}\text{C}$ - $^{13}\text{C}$ )**

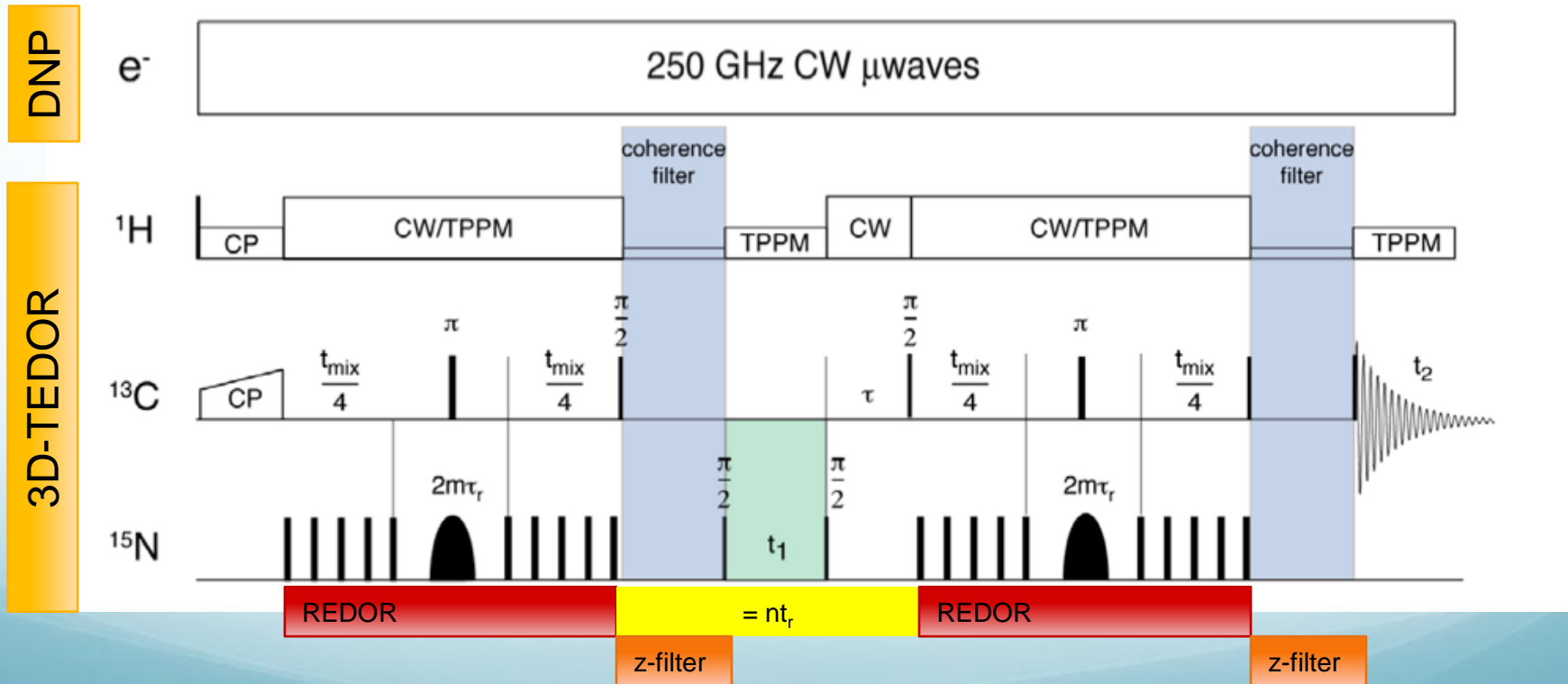
- The J-coupling imposes dephasing during the recoupling period
- The coupling can generate antiphase coherence  
Phase twisted lineshape



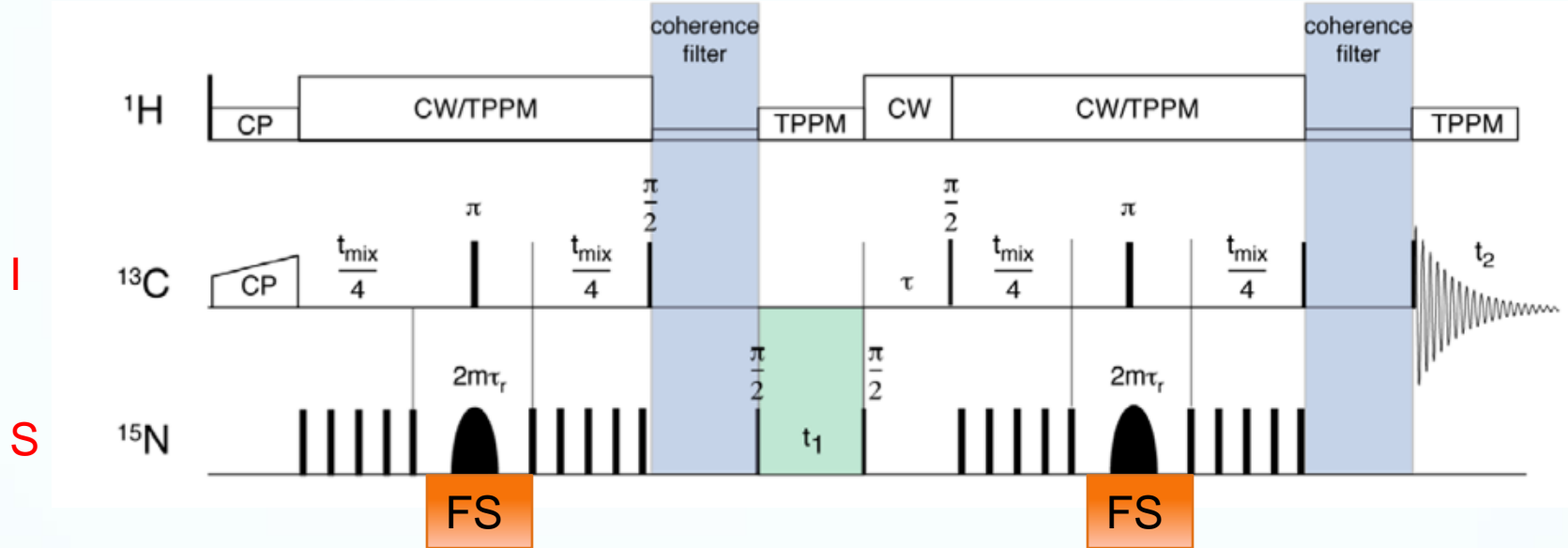
**Solution: decoupling with coherence filter**

# Methods

- 3D-TEDOR (Transfer Echo DOuble Resonance)
- Coherence filter (z-filter)
- Frequency-Selective (FS)
- DNP



# 3D TEDOR pulse sequence

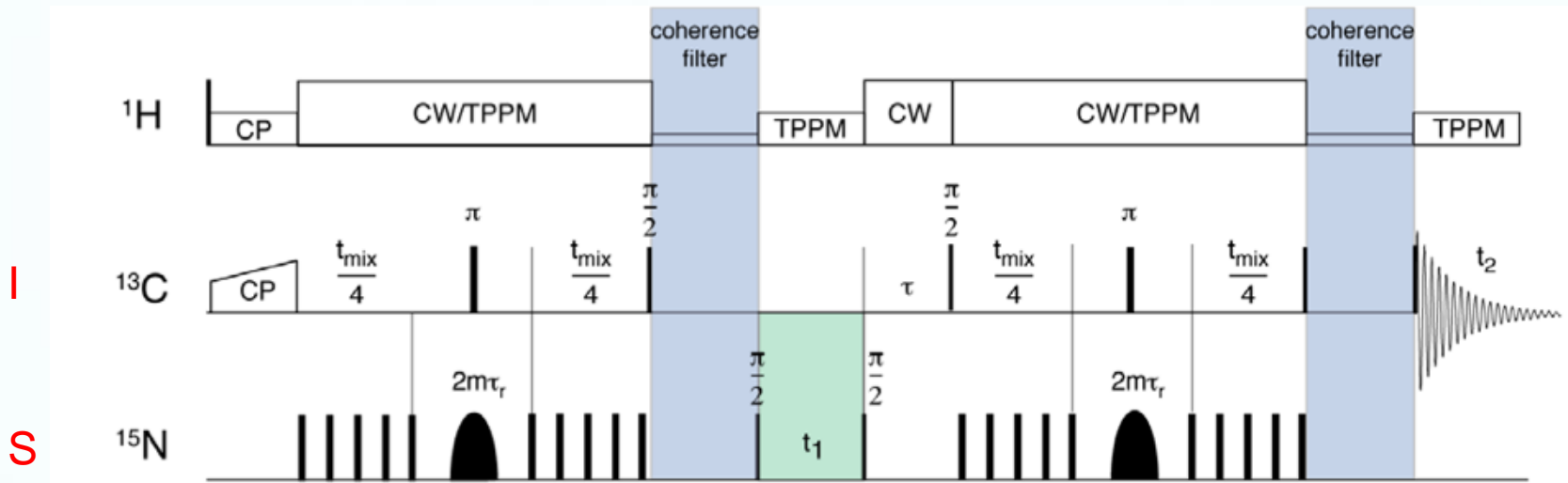


$$\begin{aligned}
 I_x &\xrightarrow{\text{REDOR}} 2I_y S_z \sin(\omega t_{\text{mix}}/2) \\
 &\xrightarrow{90_x(I)90_y(S)} -2I_z S_y \sin(\omega t_{\text{mix}}/2) \\
 &\xrightarrow{t_1} -2I_z S_y \sin(\omega t_{\text{mix}}/2) e^{i\Omega_s t_1} \\
 &\xrightarrow{90_x(I)90_{xy}(S)} -2I_y S_z \sin(\omega t_{\text{mix}}/2) e^{i\Omega_s t_1} \\
 &\xrightarrow{\text{REDOR}} I_x \sin^2(\omega t_{\text{mix}}/2) e^{i\Omega_s t_1} e^{i\Omega_I t_2}
 \end{aligned}$$

- Cross polarization
- 1<sup>st</sup> REDOR ( $t_m/2$ )
- 1<sup>st</sup> pair of  $\pi/2$  pulses
- 2<sup>nd</sup> pair of  $\pi/2$  pulses
- 2<sup>nd</sup> REDOR

$\omega$ = effective dipolar coupling,  $\Omega$ = isotropic CS

# 3D TEDOR : Spin dynamics



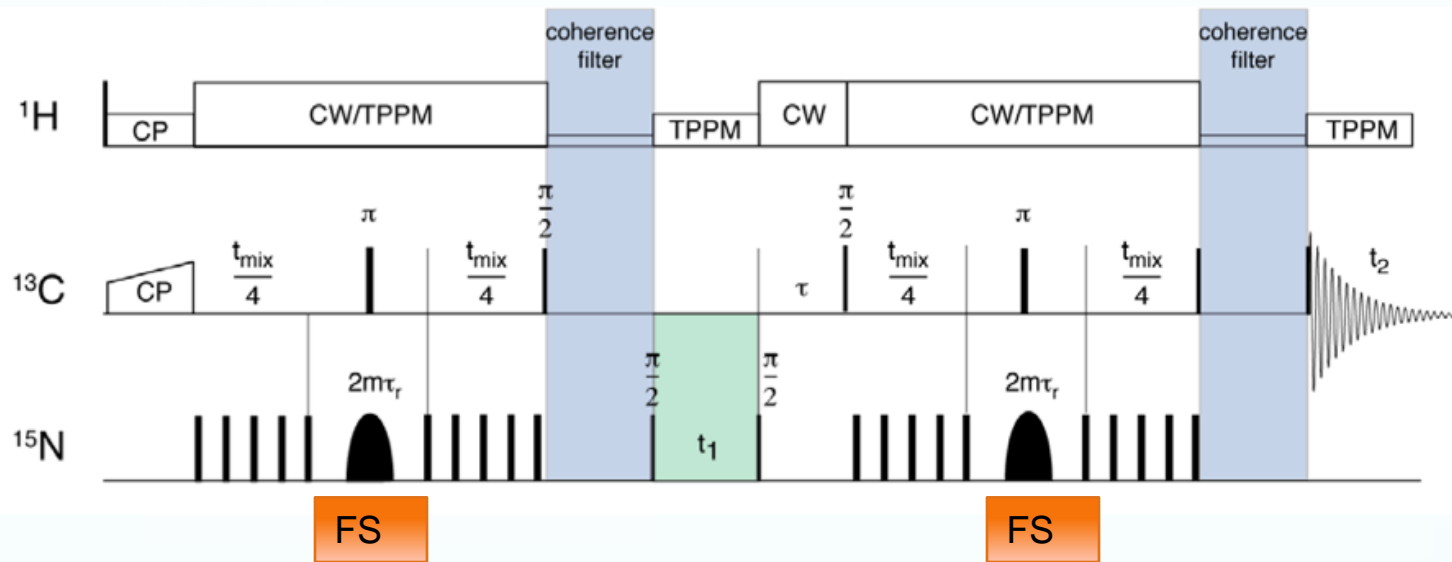
During evolution period ( $t_1$  and  $t_2$ ):

$$H = \sum_k \Omega_{Ik} I_{kz} + \sum_k \Omega_{Sk} S_{kz} + \sum_{j < k} \pi J_{jk} 2I_{jz} I_{kz}$$

During mixing period ( $t_{\text{mix}}$ ):

$$H = \sum_{j,k} \omega_{jk} 2I_{jz} S_{kz} + \sum_{j < k} \pi J_{jk} 2I_{jz} I_{kz}$$

# FS-TEDOR



Applying frequency selective pulses centered in the REDOR period

Only the nuclei within the bandwidth contribute to spin dynamics during REDOR mixing periods

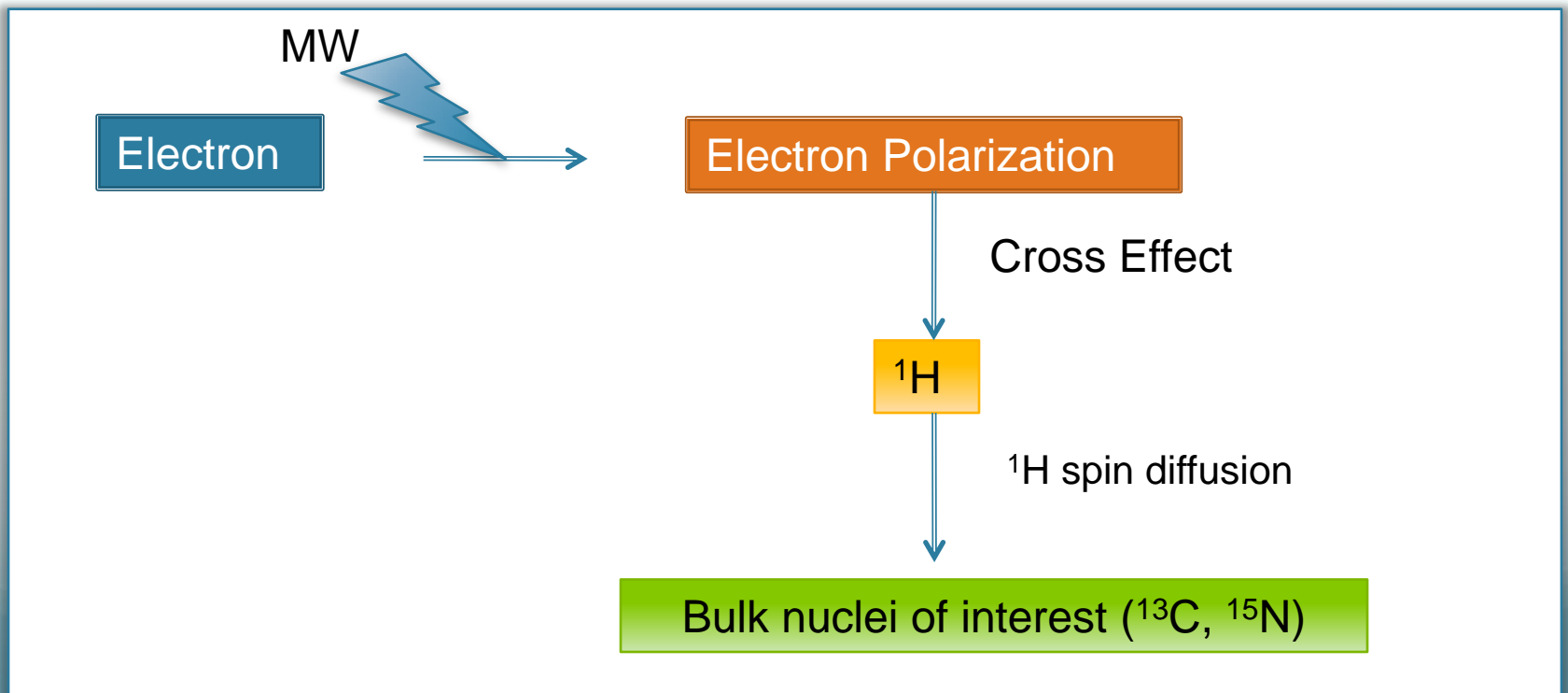


# High field DNP

DNP : Polarization transfer from electron to nuclei

Sensitivity Enhancement

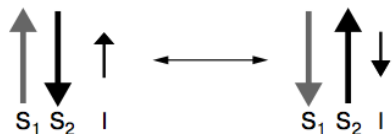
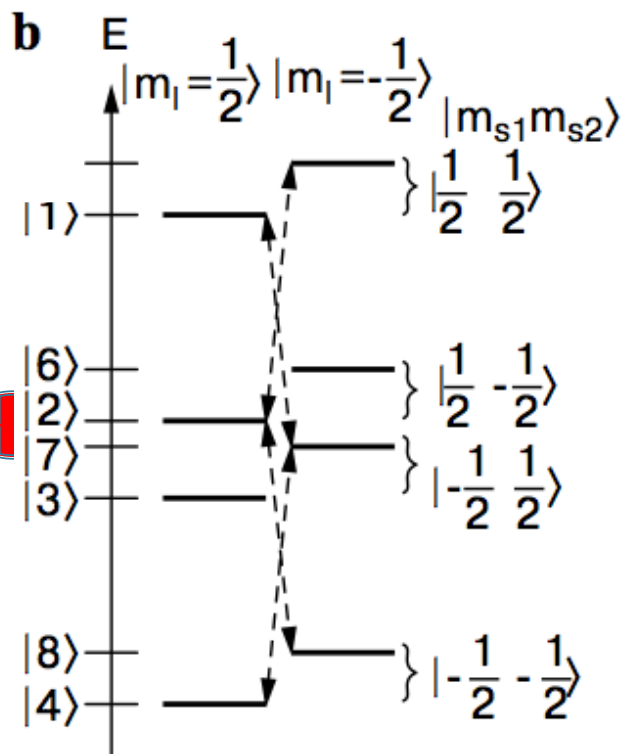
$$Y_e/Y_{1H} \approx 660$$



# High field DNP: Mechanism

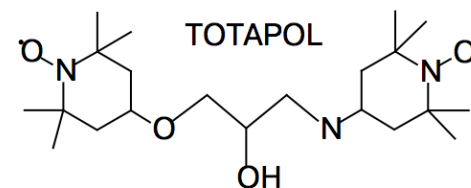
Cross Effect:

$$\delta > \omega_n$$



energy-conserved flip flop process

Biradical:



glass forming solvent

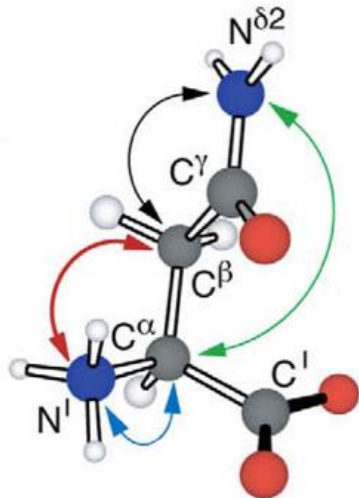
deuterated Glycerol +  
D<sub>2</sub>O + H<sub>2</sub>O

# Investigated system

## FS-TEDOR

### U- $^{13}\text{C}$ , $^{15}\text{N}$ -Asparagine

- Crystalline solid
- Uniformly labeled
- Two  $^{15}\text{N}$
- Four  $^{13}\text{C}$



## FS-TEDOR + DNP

### [U- $^{13}\text{C}$ , $^{15}\text{N}$ ]-bacteriorhodopsin

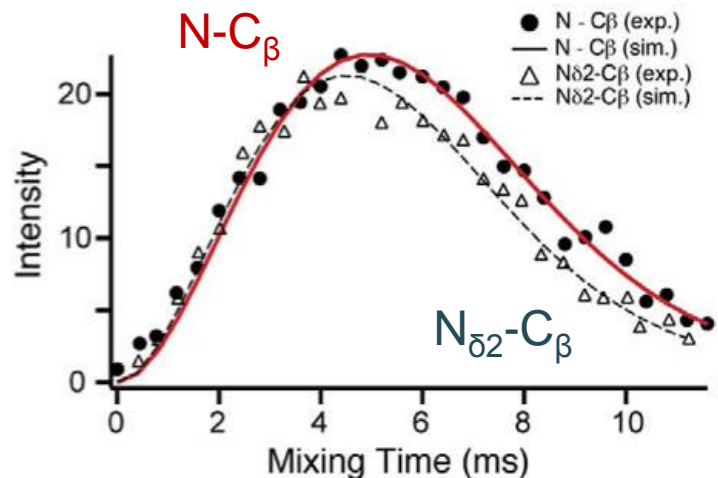
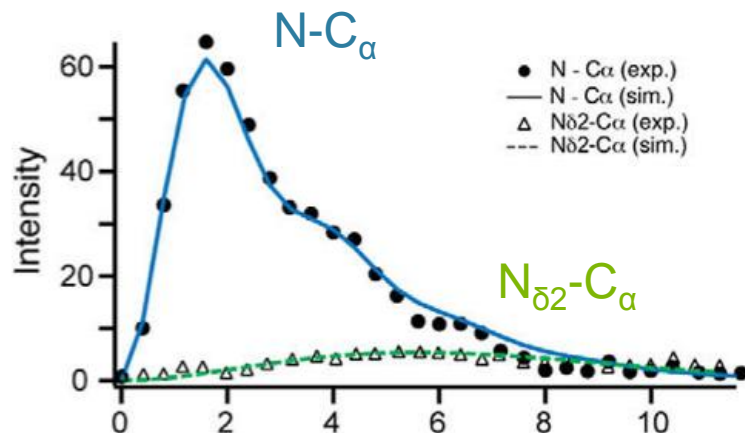
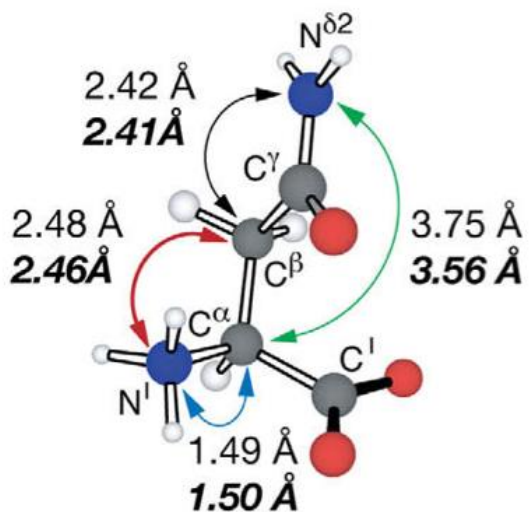
- Membrane protein
- produced by *Halobacterium Salinarum*
- Light-driven ion pump
- 26.6 kDa



# FS-TEDOR on U-[<sup>13</sup>C,<sup>15</sup>N]-Asparagine

Experiment condition:

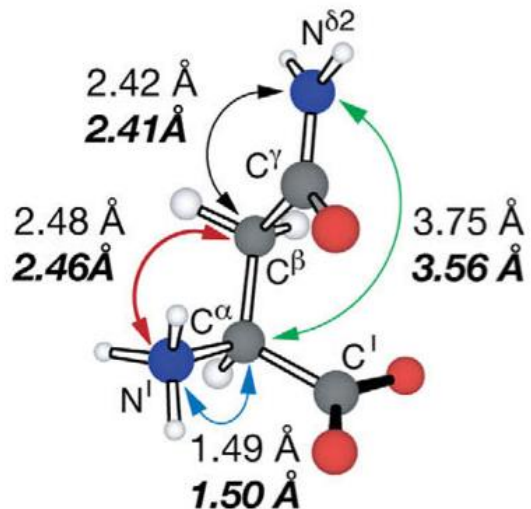
- 500 MHz(<sup>1</sup>H)
- 100 kHz TPPM
- 1ms Gaussian refocusing pulse
  - Backbone
  - Side chain
- MAS 10 kHz
- 50 kHz REDOR pulses
- 10% diluted



$$D = \left( \frac{\mu_0}{4\pi} \right) \frac{\gamma_I \gamma_S \hbar}{2\pi r^3}$$

# Distance measurements by FS-TEDOR as compared to other techniques

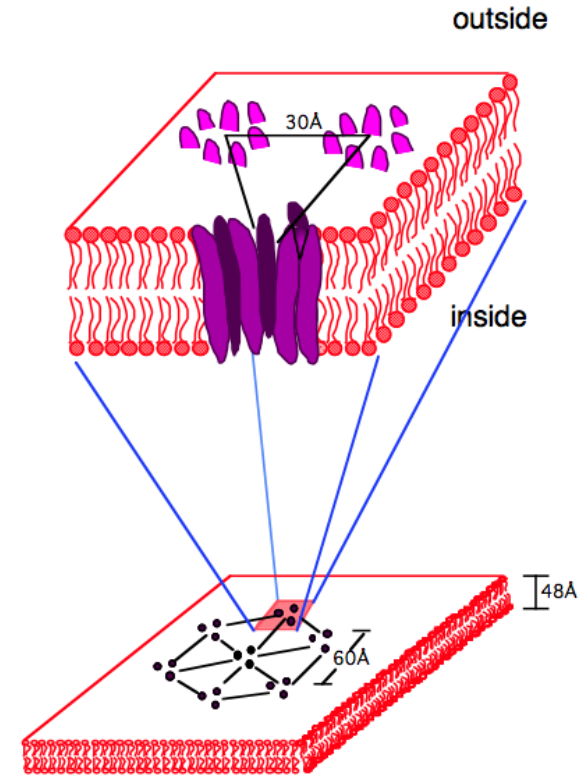
	FS-TEDOR (Å)	FS-REDOR (Å)	Neutron diffraction (Å)
N-C <sup>α</sup>	1.50	1.50	1.49
N <sup>δ2</sup> -C <sup>α</sup>	3.56	3.58	3.75
N-C <sup>β</sup>	2.46	2.49	2.48
N <sup>δ2</sup> -C <sup>β</sup>	2.41	2.44	2.42



The results are in good agreement with other methods

# [U-<sup>13</sup>C, <sup>15</sup>N]-bacteriorhodopsin

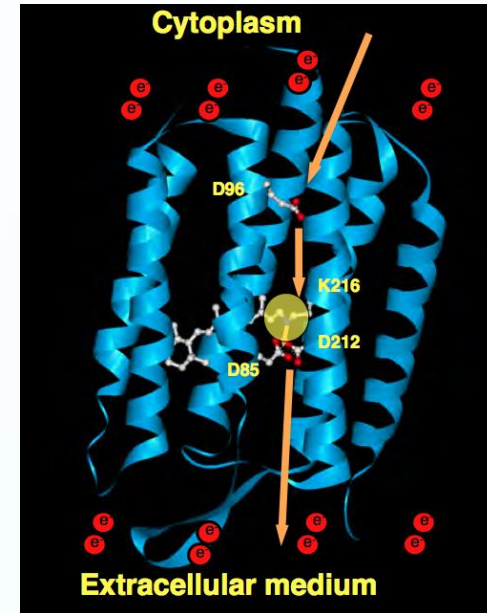
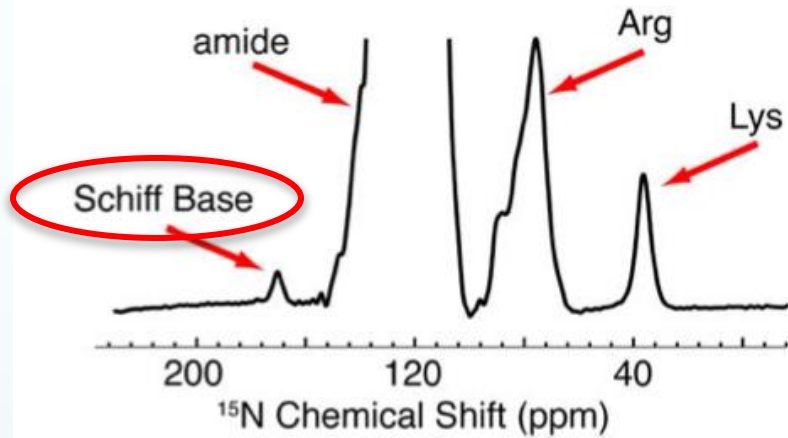
- Light-driven ion pump
- Seven transmembrane helixs
- Homotrimer
- Homotrimers aggregate to form a purple membarane
- The retinal chromophore is attached via a Schiff base linkage to Lys216
- Arg82 is part of the complex counterion



Ref [6]

# Bacteriorhodopsin: 1D DNP enhanced $^{15}\text{N}$ spectrum

$\epsilon = 43$  at 200 K  
 $\epsilon = 90$  at 200 K

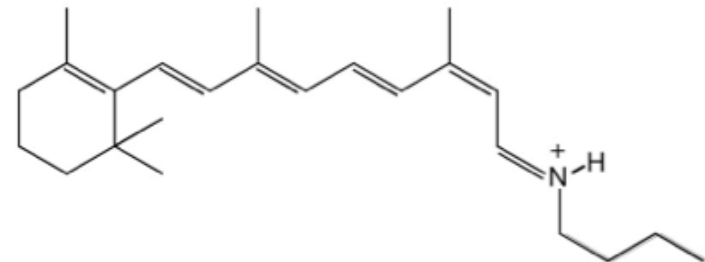


Ref [6]

Excellent S/N!

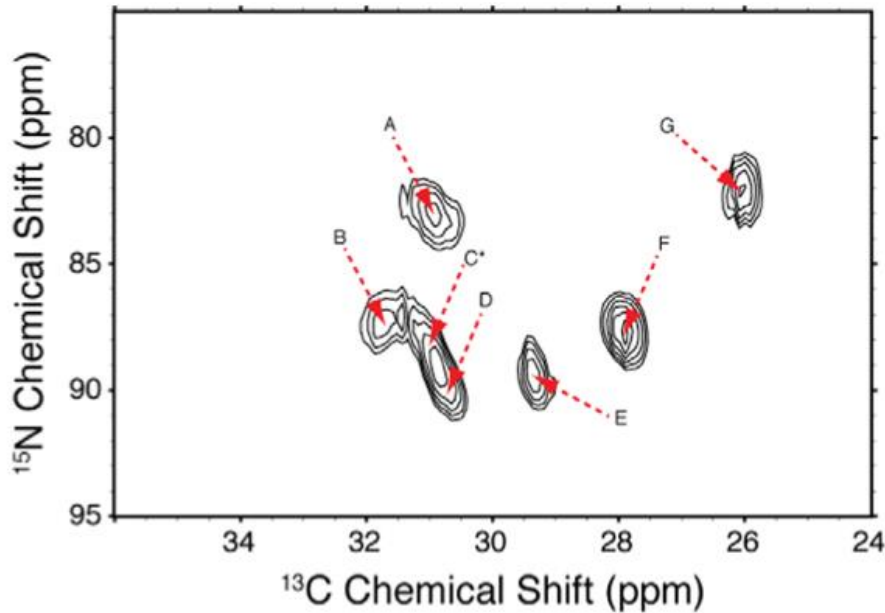
MAS-NMR  
CW MW irradiation at 250GHz  
at 90K  
Polarizing agent: TOTAPOL

K,L (protonated)



Ref [5]

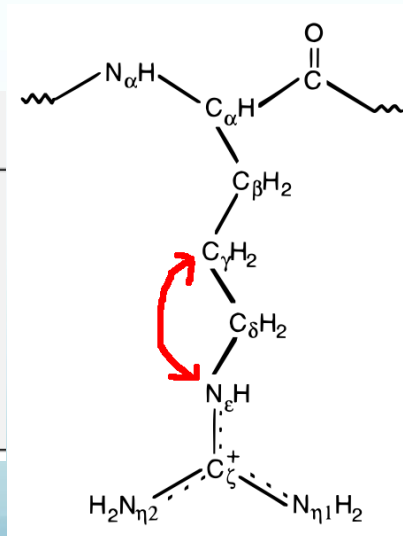
# Bacteriorhodopsin: 2D $^{15}\text{N}_\gamma$ - $^{13}\text{C}_\epsilon$ selective correlation spectra



## 7 Arg- $^{13}\text{C}_\gamma$ - $^{15}\text{N}_\epsilon$ of bR

- MAS 7.576 kHz, 380 MHz  $^1\text{H}$
- CW MW irradiation at 250GHz
- Temperature = 90K
- Polarizing agent: TOTAPOL
- 1.98 ms Gaussian refocusing pulse (selective pulse center at 90 ppm)
- REDOR mixing time 4 ms (favorable 2.4 -2.5 Å)

Peak	$^{15}\text{N}$ chemical shift (ppm)	$^{13}\text{C}$ chemical shift (ppm)	Peak volume (arbitrary units)
A	82.9	30.9	2.9
B	87.3	31.7	2.7
C <sup>a</sup>	87.8	31.0	2.7
D	89.9	30.7	3.1
E	89.4	29.3	3.0
F	87.6	27.9	3.5
G	82.1	26.0	3.1



Arg $^{15}\text{N}_\epsilon$ : 90±19 ppm

Arg $^{15}\text{N}_{\eta_1}$ : 75±14 ppm

Arg $^{15}\text{N}_{\eta_2}$ : 75±16 ppm

Arg- $^{13}\text{C}_\gamma$ : 27±2 ppm

Arg- $^{13}\text{C}_\delta$ : 43.2±2 ppm



# Conclusion

- FS-TEDOR can be used quantitatively and qualitatively for  $^{15}\text{N}$ - $^{13}\text{C}$  correlation spectroscopy in crystalline solids and membrane proteins.
- Six of seven Arg- $^{13}\text{C}_\gamma$ - $^{15}\text{N}_\epsilon$  correlation spectra have been resolved using DNP.
- The method may be contemplated in lieu of specific isotopic labelling or suppression to simplify the spin dynamics.

# References

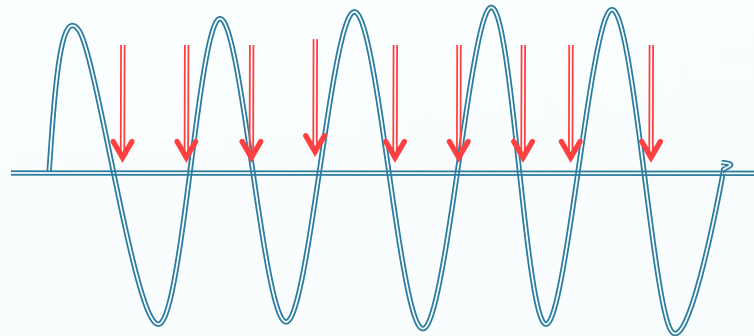
1. V. S. Bajaj, M. L. Mark-Jurkauskas, M. Belenky, J. Herzfeld, R. G. Griffin, *J. Magn. Reson.* **2010**, *202*, 9-13.
2. C. P. Jeroniec, C. Filip, R. G. Griffin, *J. Am. Chem. Soc.* **2002**, *124*, 10728-10742.
3. A. B. Barnes, G. D. Paepe, P. C. A. van der Wel, K.-N. Hu, C.-G. Joo, V.S. Bajaj, M. L. Mak-Jurkauskas, J.R. Sirigiri, J. Herzfeld, R. J. Temkin, R. G. Griffin, *Appl. Magn. Resonan.* **2008**, *34*, 237-263.
4. A. T. Petkova, J. G. Hu, M. Bizounok, M. Simpson, R. G. Griffin, J. Herzfeld, *Biochemistry* **1999**, *38*, 1562-1572.
5. V. S. Bajaj, M. L. Mark-Jurkauskas, M. Belenky, J. Hrzfeld, R. G. Griffin, *Proc. Natl. Acad. Sci. USA*, **2009**, *106*, 9244-9249.
6. Lecture (Francis Bitter Magnet Laboratory and Department of Chemistry Massachusetts Institute of Technology), [nmrwinterschool.com](http://nmrwinterschool.com) (accessed Sep, 17<sup>th</sup>, 2013)

Thank you very much

# Supplements

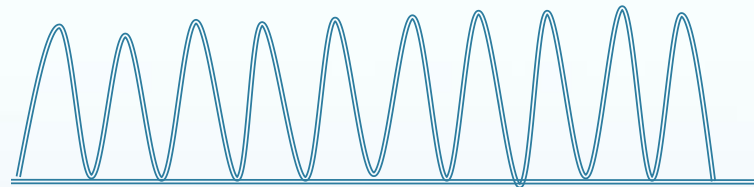
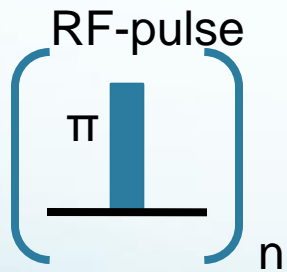
# REDOR

MAS



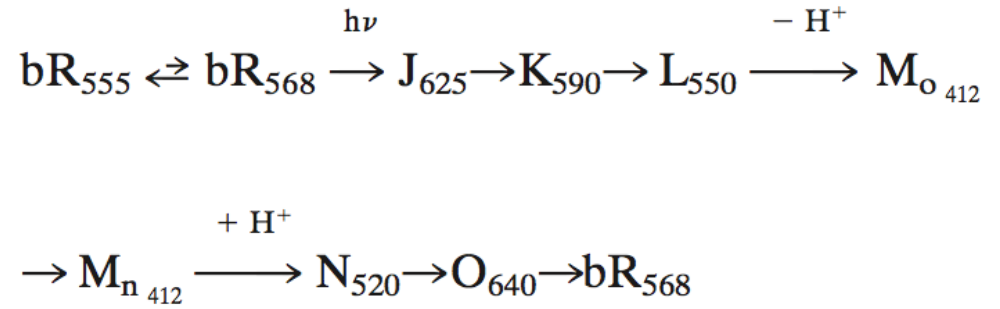
$$\omega_{\text{dip}}(t) = 0$$

REDOR

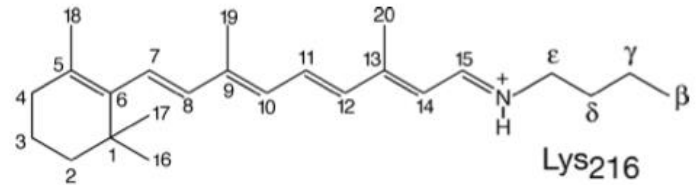


$$\omega_{\text{dip}}(t) \neq 0$$

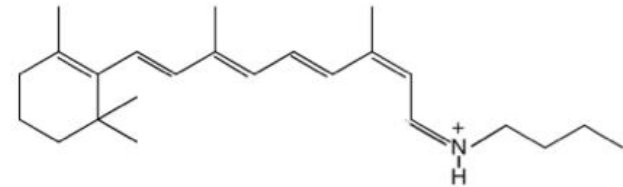
# bR: Photocycle



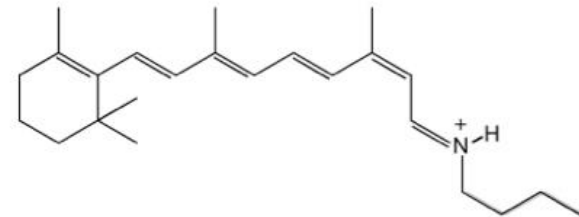
$\text{bR}_{568}$   
 13-trans, 15-anti  
 protonated



$\text{bR}_{555}$   
 13-cis, 15-syn  
 protonated



K, L  
 13-cis, 15-anti  
 protonated



$\text{M}_o$   
 13-cis, 15-anti  
 deprotonated

